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Efficient high-throughput screening by ER Ca²⁺ measurement to identify inhibitors of ryanodine receptor Ca²⁺-release channels.

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Abstract

Genetic mutations in **ryanodine** receptors (RyRs), **Ca²⁺-release channels** in the sarcoplasmic reticulum essential for muscle contractions, cause various skeletal muscle and cardiac diseases. Because the main underlying mechanism of the pathogenesis is overactive **Ca²⁺ release** by gain-of-function of the RyR channel, inhibition of RyRs is expected to be a promising treatment for these diseases. Here, to **identify inhibitors** specific to skeletal muscle type 1 RyR (RyR1), we developed a novel **high-throughput screening** (HTS) platform using time-lapse fluorescence **measurement of Ca²⁺ concentrations in the endoplasmic reticulum (ER) ([Ca²⁺]ER)**. Because expression of RyR1 carrying disease-associated mutation reduces **[Ca²⁺]ER** in HEK293 cells through **Ca²⁺ leakage from RyR1 channels**, specific drugs that inhibit RyR1 will increase **[Ca²⁺]ER** by preventing such **Ca²⁺ leakage**. RyR1 carrying R2163C mutation and R-CEPIA1er, a genetically-encoded **ER Ca²⁺ indicator**, were stably expressed in HEK293 cells and time-lapse fluorescence was measured using a FlexStation II fluorometer. False positives were effectively excluded by using cells expressing wild-type (WT) RyR1. By **screening** 1,535 compounds in a library of well-characterized drugs, we successfully identified four compounds that significantly increased **[Ca²⁺]ER**. They include dantrolene, a known RyR1 inhibitor, and three structurally-different compounds; oxolinic acid, 9-aminoacridine and alexidine. All the hit compounds, except for oxolinic acid, inhibited **[³H]ryanodine** binding of WT and mutant RyR1. Interestingly, they showed different dose-dependencies and isoform specificities. The highly quantitative nature and good correlation

with the channel activity validated this HTS platform by [**Ca²⁺**]ER measurement to explore drugs for RyR-related diseases.

KEYWORDS: Calcium; Calcium imaging; Drug discovery; **High throughput screening**; Ion channels; **Ryanodine** receptors; Skeletal muscle

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